

## SNP Analysis using dartR



# Guide to Basic Filtering

Version 1.8.3



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# Session 1: Basic Filtering

## Overview



DART Pty Ltd has already done much of the filtering of the sequences used to generate your SNPs that would normally be undertaken by researchers who generate their own ddRAD data. Here we present some other filters that you might wish to apply.

It is a good idea to run the `gl.report` functions in advance of filtering to provide a foundation for selecting thresholds.

Several filters are available to improve the quality of the data represented in your `genlight` object. The basic ones are:

<code>gl &lt;- gl.filter.reproducibility()</code>	filter out loci for which the reproducibility (strictly repeatability) is less than a specified threshold, say threshold = 0.99
<code>gl &lt;- gl.filter.callrate()</code>	filter out loci or individuals for which the call rate (rate of non-missing values) is less than a specified threshold, say threshold = 0.95
<code>gl &lt;- gl.filter.monomorphs()</code>	filter out monomorphic loci and loci that are scored all NA
<code>gl &lt;- gl.filter.secondaries()</code>	filter out SNPs that share a sequence tag, except one retained at random
<code>gl &lt;- gl.filter.hamming()</code>	filter out loci that differ from each other by less than a specified number of base pairs
<code>gl &lt;- gl.filter.rdepth()</code>	filter out loci with exceptionally low or high read depth (coverage)
<code>gl &lt;- gl.filter.taglength()</code>	filter out loci for which the tag length is less than a threshold
<code>gl &lt;- gl.filter.overshoot()</code>	filter out loci where the SNP location lies outside the trimmed sequence tag

The order of filtering can be important and requires some thought. Filtering on call rate by individual before filtering on call rate by locus or choosing the alternative order will depend on the weight placed on losing individuals versus losing loci, for example.

### Example: Filtering on reproducibility

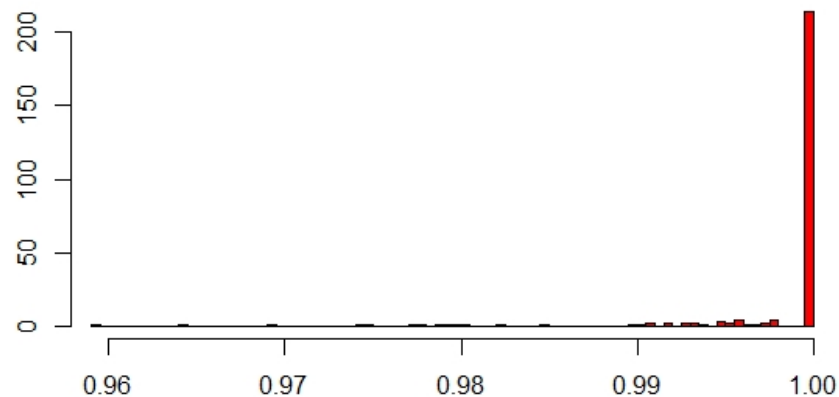
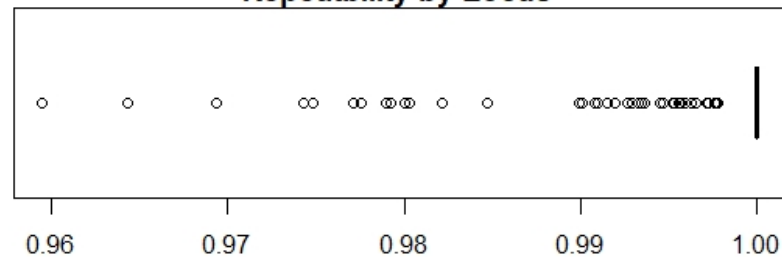
First, we should examine the distribution of reproducibility measures (`RepAvg`) in our dataset.

```
gl.report.reproducibility(testset.gl,plot=TRUE)
```

```
Starting gl.report.reproducibility
Processing a SNP dataset
No. of loci = 255
No. of individuals = 250
Minimum repeatability: 0.96
Maximum repeatability: 1
Mean repeatability: 0.998
Boxplot adjusted to account for skewness
```

```
Completed: gl.report.reproducibility
Threshold Retained Percent Filtered Percent
1 1.0000000 214 83.9 41 16.1
2 0.9979729 214 83.9 41 16.1
3 0.9959459 222 87.1 33 12.9
4 0.9939189 231 90.6 24 9.4
5 0.9918918 237 92.9 18 7.1
6 0.9898647 242 94.9 13 5.1
7 0.9878377 242 94.9 13 5.1
8 0.9858107 242 94.9 13 5.1
9 0.9837836 243 95.3 12 4.7
10 0.9817565 244 95.7 11 4.3
11 0.9797295 246 96.5 9 3.5
12 0.9777025 248 97.3 7 2.7
13 0.9756754 250 98.0 5 2.0
14 0.9736483 252 98.8 3 1.2
15 0.9716213 252 98.8 3 1.2
16 0.9695942 252 98.8 3 1.2
17 0.9675672 253 99.2 2 0.8
18 0.9655401 253 99.2 2 0.8
19 0.9635131 254 99.6 1 0.4
20 0.9614860 254 99.6 1 0.4
21 0.9594590 255 100.0 0 0.0
```

**SNP data (DARTSeq)  
Repeatability by Locus**



Clearly, with a minimum repeatability of 0.96 and a maximum of 1 across loci, we can be fairly stringent in our choice of a threshold. A value of 0.99 will not result in the loss of much data.

We now filter on that basis.

```
gl <- gl.filter.reproducibility( testset.gl,
                               threshold=0.99, verbose = 3)
```

```
Starting gl.filter.reproducibility
Processing a SNP dataset
Identifying loci with repeatability below : 0.99
Removing loci with repeatability less than 0.99
```

```
Summary of filtered dataset
Retaining loci with repeatability >= 0.99
Original no. of loci: 255
No. of loci discarded: 14
No. of loci retained: 241
No. of individuals: 250
No. of populations: 30
```

```
Completed: gl.filter.reproducibility
```

Only 14 loci out of 255 were deleted.

It is wise not to filter too heavily on reproducibility. A threshold of 1 is often tempting but can result in some bias being introduced.



### Exercises

1. Just in case you have accidentally modified the genlight object `gl`, recreate it by copying it from `testset.gl`.
2. Filter `gl` using the filters listed above. Request a report first to inform your choice of threshold. Be sure to set `plot=TRUE` to examine the distribution of each parameter and optionally `smearplot=TRUE` to examine the smear plot.

## Recalculating locus metadata after filtering

Remember, the locus metrics are no longer valid if individuals or populations are deleted from the dataset. For example, if you filter out a population for which the individuals have particularly bad call rates, then the call rate parameter held in the locus metrics will no longer be accurate. It will need to be recalculated. This is true of many of the locus metrics.

So, after filtering your data, it is wise to recalculate the locus metrics with

```
gl <- gl.recalc.metrics(gl)
```



Try this for yourself on genlight object `gl` after filtering or on your own data

Similarly, when filtering has resulted in removal of some individuals or populations, variation at several loci may be lost. Some loci may even be scored as missing across all individuals. You may wish to remove these monomorphic loci from your dataset with

```
gl <- gl.filter.monomorphs(gl)
```



Try this for yourself on genlight object `gl` after filtering or on your own data

## Where have we come?

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The above Session was designed to give you an overview of the scripts in dartR for filtering your data. Having completed this Session, you should now be able to:

- Filter on call rate, repeatability, secondaries, hamming distance, and minor allele frequency.
- Recalculate locus metrics after deleting individuals or populations as part of the filtering process.
- Filter out resultant monomorphic loci.

## Further reading

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Jombart T. and Caitlin Collins, C. (2015). Analysing genome-wide SNP data using adegenet 2.0.0. <http://adegenet.r-forge.r-project.org/files/tutorial-genomics.pdf>

Gruber, B., Unmack, P.J., Berry, O. and Georges, A. 2018. dartR: an R package to facilitate analysis of SNP data generated from reduced representation genome sequencing. *Molecular Ecology Resources*, 18:691–699



Ende